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MODULATION OF CULTURED LEYDIG CELL STEROIDOGENIC ACTIVITY BY SERTOLI CELL SECRETED FACTORS.

A.M.MORERA, M.BENAHMED, M.A.CHAUVIN, C.GRENOT and E.de PERETTI.
INSERM U34 and U162 Hopital Debrousse 69322 LYON FRANCE

Using a model of coculture of immature porcine Sertoli and Leydig cells, we have shown that FSH enhances Leydig cell activity via Sertoli cells. Conditioned medium (CM) from FSH-stimulated Sertoli cells (FSH-CM) mimics the effect of FSH-stimulated Sertoli cells, suggesting that Sertoli cells act via diffusible factors. Here, we have investigated the effect of such factors on Leydig cell activity by two parameters: 125-I hCG binding and steroid production (testosterone, DHAS and Pregnenolone sulfate). We have compared the activity of Leydig cells cultured in FSH-CM to CM from control Sertoli cells (C-CM). An increase of 125-I hCG binding and basal and hCG-stimulated steroid production is observed in Leydig cells cultured in FSH-CM versus C-CM. Scatchard analysis of hCG receptors show an increase of hCG binding sites without modification of hCG receptor affinity. Dose-response curve to hCG shows that maximal testosterone response is increased whereas the ED 50 is not modified. The steroidogenic response to hCG, PGE₂, 8Br cAMP and forskolin is increased. The presence of LDL-cholesterol in the culture medium potentiates the steroidogenic effect of FSH-CM. These results show that FSH-dependent factors from Sertoli cells modulate Leydig cell activity by several mechanisms: 1) hCG binding sites number, 2) exogenous cholesterol utilization, 3) response to cAMP stimulation. Precise sites of action of these factors on steroid biosynthesis are under investigation.

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CONFORMATIONAL CHANGES INDUCED BY STEROID HORMONES BINDING TO SERUM ALBUMIN: A KINETIC STUDY

Gianfranco GIRAUDI - Department of Analytical Chemistry, University of Torino, Torino

The kinetics of the association reaction between bovine serum albumin (BSA) and estradiol (E₂), progesterone (Pg), testosterone (T), 5 α -dihydrotestosterone (DHT) and androstenedione (A) was studied at 9.5^oC, pH 7.4, 0.15 M ionic strength, by a stopped-flow spectrophotometric technique by following the time course of the fluorescence quenching of the BSA upon steroid binding. For all steroids examined two first-order steps were observed: a fast fluorescence quenching followed by a much slower one. With increasing concentrations of excess steroid, the first-order rate constants showed an enzyme-like saturation kinetics for the first step, but were constant for the second step. This trend provides direct evidence of a complex reaction mechanism of the type: $P + S \xrightleftharpoons{K_1} C \xrightleftharpoons{K_2} D \xrightleftharpoons{K_3} E$, where a very fast formation of an intermediate complex (C) is followed by a minor isomerization step (2) and then by a much slower, more extensive rearrangement (3). The derived equilibrium and kinetic constants (K₁ (1/mol), K₂, K_{eq} (1/mol), k₂ (1/s), and k₋₂ (1/s), respectively) for some steroids are: Pg) 5.8·10⁴, 2.6, 2.1·10⁵, 8.4, 3.2 E₂) 5.3·10⁴, 2.5, 1.9·10⁵, 12.5, 5 DHT) 3.6·10⁴, 2.0, 1.1·10⁵, 20, 10. The first-order rate constants of the slower step (3) are almost independent on steroid structure. The rise of K_{eq} together with the lowering of the forward and reverse rate constants with increasing K₂, shows clearly how a tight binding in the isomerization step can affect both the kinetics and the equilibrium of steroid-protein interactions.